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# Development of Dual-Acting Agents for Thromboxane Receptor Antagonism and Thromboxane Synthase Inhibition—I. Synthesis, Structure–Activity Relationship, and Evaluation of Substituted ω-Phenyl-ω-(3-Pyridyl)Alkenoic Acids

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Abstract—A series of arylsulfonamido-substituted  $\omega$ -phenyl- $\omega$ -(3-pyridyl)alkenoic acids were synthesized and evaluated in vitro for their ability to act as both a thromboxane  $A_2$  receptor antagonist (TRA) and thromboxane synthase inhibitor (TSI). Variations of alkenoic acid chain length, olefin geometry, substituent effect on the benzenesulfonamido group, and conformational flexibility of the substituted arylsulfonamido group were examined. Among the various substituents, iodo-substitution gave the most potent compound. Conformational flexibility between the arylsulfonamido group and the phenyl ring attached to the alkenoic acid side chain significantly enhanced the dual activities. The compound (E)-21c was identified as the most potent TRA/TSI (TRA:  $K_d = 53$  nM; TSI: IC $_{50} = 23$  nM) in the series studied. The compounds 9c and 10c have indicated that these series of compounds are orally active and are specific TSIs as exhibited by the so-called 'shunt' effect on prostacyclin synthesis in vitro.

#### Introduction

Modulation of either the synthesis or the activity of arachidonic acid metabolites continues to be an area that attracts substantial research interest in potential therapeutic agents for various circulatory disorders. 1-4 An appropriate balance between the biosynthesis of thromboxane A2 (TXA<sub>2</sub>, 1) and prostacyclin (PGI<sub>2</sub>, 2), two extremely unstable metabolites of arachidonic acid with opposing activities, is considered an important factor for maintaining a normal hemodynamic status. In particular, TXA2, being an extremely potent platelet-aggregating and vasoconstricting agent, plays a major role in the pathogenesis of certain vascular and renal disorders.<sup>5</sup> Thromboxane synthase inhibitors (TSIs) and thromboxane receptor antagonists (TRAs) have been developed to treat these disorders. 2-4 While the efficacy of TRAs has yet to be established, TSIs have performed poorly in clinical trials.<sup>6</sup> The lack of efficacy of TSIs has been ascribed to PGH<sub>2</sub> (3) which accumulates from inhibition of thromboxane synthase and whose agonist activity nullifies the benefits of reducing TXA<sub>2</sub> levels. Theoretical arguments have been made to support the potentially superior antithrombotic efficacy of using a combined TRA/TSI over either class of agent alone or aspirin.<sup>7-10</sup> Thromboxane synthase inhibitory action of such an agent would prevent the biosynthesis of TXA2 while the accumulated PGH2 would be redirected and converted to

beneficial prostaglandins such as PGI<sub>2</sub> (prostacyclin) and PGD<sub>2</sub>. This so-called 'shunt' effect is not possible by the use of a TRA or aspirin which inhibits prostacyclin production. The thromboxane receptor antagonism of the combined agent, at the same time, would antagonize the action of PGH<sub>2</sub>. We and others<sup>11-14</sup> have undertaken a program to develop compounds which contain both TRA and TSI activities in a single chemical entity. Such compounds would minimize and overcome the limitation of aspirin or a TRA or TSI alone and would solve the impracticality of using two drugs with potentially different pharmacokinetics for therapeutic purposes.

In this paper, we describe the design, synthesis, structure and activity relationship (SAR), and in vitro and in vivo pharmacology of a series of compounds which possess TRA/TSI activity.

#### **Compound Design**

Our strategy to identify compounds which possess dual TRA and TSI was based upon a design of hybrid structures by incorporating the basic pharmacophore for each activity. The essential structural features of a TSI are a basic nitrogen atom of a 3-substituted pyridine or a 1-substituted imidazole ring and a carboxylic acid functionality separated by a distance of 8.5–10 Å. 15 The pharmacophores for TRA vary widely in structural type, of which a combination of aryl sulfonamide and carboxylic acid is well known. In the process of our program development, the compound 4 (LY280978) was identified as a potent TRA. 16 Design of the target molecule I which

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incorporates a pyridyl ring into 4 to hopefully induce a balanced combined TRA/TSI activity appeared promising. Examples such as 5 (CV-4151)<sup>15</sup> and 6 (R68070)<sup>28</sup> have been reported to possess a combined TRA/TSI activity albeit with modest TRA.<sup>4,9</sup>

# Chemistry

The initial target molecules 9 and 10 were synthesized via a Wittig reaction of the benzenesulfonamido-substituted phenyl pyridyl ketones 7 and 8, respectively, as shown in Scheme I. The ketones 7 and 8 were prepared from commercially available 3- or 4-bromoaniline in three steps: (1) sulfonamide formation (PhSO<sub>2</sub>Cl, pyridine, 0 °C, 98 %); (2) phenyl(pyridyl)carbinol formation (n-BuLi, THF, -78 °C; 3-pyridinecarboxaldehyde, 59-60 %); and (3) oxidation of the carbinols to the ketones by any one of the conventional oxidation methods such as Swern, Jones, or

preferably MnO<sub>2</sub> oxidation (MnO<sub>2</sub>, 1,4-dioxane, 70 °C overnight, 96 %). The Wittig reaction of these ketones with ( $\omega$ -carboxyalkyl)triphenylphosphonium bromide<sup>15</sup> in the presence of NaHMDS, THF, -78 °C (34-55 %) or *t*-BuOK, THF, -10 ~ 0 °C (88-100 %) yielded mixtures of (*E*)- and (*Z*)-alkenoic acids 9 and 10, predominantly as the (*E*)-isomers, <sup>17</sup> as shown in Table 1.

In order to assess the effect of the double bond geometry of the alkenoic acids and substituent effect on the sulfonamide phenyl ring, the chemistry was modified to separate the two geometric isomers as shown in Scheme II. The nitrophenyl pyridyl ketones 11 and 12 were obtained in two steps from 3-bromopyridine: (1) nitrophenyl-(pyridyl)carbinol formation (*n*-BuLi, Et<sub>2</sub>O, -78 °C; nitrobenzaldehyde, 35–54 %); and (2) MnO<sub>2</sub> or Swern oxidation of the carbinols to the ketones (94–98 %). The ketones were then subjected to a Wittig reaction followed

Scheme I. a) PhSO<sub>2</sub>Cl, pyr.; b) n-BuLi, THF, -78 °C; 3-pyridinecarboxaldehyde; c) MnO<sub>2</sub>, dioxane, 70 °C; d) Br $^-$ Ph<sub>3</sub>P $^+$ (CH<sub>2</sub>)  $_{n+1}$ CO<sub>2</sub>H, & BuOK, THF, -10-0 °C.

Table 1. In vitro activities of compounds 9 and 10

| N OH                 | N Company                  |
|----------------------|----------------------------|
| NHSO <sub>2</sub> Ph | 10a-d NHSO,Ph              |
| 74-1                 | 10a-d NHSO <sub>2</sub> Ph |

| Compounds   | n | <u>E/Z</u> a   | TRA<br><u>Kđ (µM)</u> b | TSI<br><u>IC50 (nM)</u> b |
|-------------|---|----------------|-------------------------|---------------------------|
| 9a          | 2 | 90 : 10        | $4.0 \pm 0.3$           | 8,000                     |
| 9b          | 3 | 77 : <b>23</b> | $3.9 \pm 0.3$           | 2,000                     |
| 9c          | 4 | 84 : 16        | $2.8\pm0.2$             | 230                       |
| 9d          | 5 | 100:0          | $2.7 \pm 0.2$           | 180                       |
| 9e          | 6 | 96:4           | $5.8 \pm 0.5$           | 400                       |
| <b>9</b> f  | 7 | 73:27          | $2.7 \pm 0.4$           | 93                        |
| 10a         | 3 | 98:2           | $1.2 \pm 0.15$          | 2,100                     |
| 1 <b>0b</b> | 4 | 97:3           | $1.1 \pm 0.20$          | 2,200                     |
| 10c         | 5 | 93:7           | $0.324 \pm 0.031$       | 130                       |
| 10d         | 6 | 96 : 4         | $2.3 \pm 0.40$          | 140                       |
| 6           |   |                | 8.0 ± 0.35              | 14.7±4.9                  |

"The (E/Z) ratio was determined by HPLC analysis.

by esterification (( $\omega$ -carboxyalkyl)triphenylphosphonium bromide, t-BuOK, THF, 0 °C; CH<sub>2</sub>N<sub>2</sub>, Et<sub>2</sub>O-THF, 0 °C, 45-57 %). In this case, the (Z)-isomers (the less polar material) were the predominant products in all cases (E/Z=1/~3 for meta-series; E/Z=1/~5 for para-series). The (E/Z)-mixture of 13 and 14 were separated by chromatography at this stage. Each isomer was then subjected to the following conditions to furnish the desired products 15 and 16: (1) amine formation (NiCl<sub>2</sub>·6H<sub>2</sub>O,

NaBH<sub>4</sub>, MeOH, 0 °C, <sup>18</sup> 65–88 %); (2) sulfonamide formation followed by hydrolysis (*p*-X-PhSO<sub>2</sub>Cl, pyr., 0 °C to rt; 1.0 N NaOH, THF–MeOH; H<sup>+</sup> 40–92 %).

Scheme III describes the synthetic route to the series 21 and 22. These compounds were synthesized in 11 steps from commercially available 1,3- or 1,4-benzenedimethanol. Benzenedimethanol was mono-protected with t-butyldimethylsilyl chloride (TBSCl) by a standard method (36-48 %). The mono-protected diol was oxidized with MnO<sub>2</sub> in THF at reflux to afford the aldehyde (83–86 %), which was then treated with 3-lithiopyridine (generated by the reaction of 3-bromopyridine with n-BuLi at -78 °C in Et<sub>2</sub>O). The pyridylcarbinol thus obtained (79-89 %) was oxidized with MnO<sub>2</sub> to the corresponding ketones 17 and 18 (96-97 %). These ketones were then subjected to a Wittig reaction with an (ω-carboxyalkyl)triphenylphosphonium bromide and t-BuOK in THF at 0 °C. The two geometrical (E)- and (Z)-isomers obtained could be separated by chromatography at this stage or in a subsequent step. In all cases the (Z)-isomer (less polar material) was the predominant Wittig reaction product  $(E/Z = \sim 1 / 2.5 \text{ for the } m\text{-substituted phenyl derivatives};$ E/Z = -1/5 for the p-substituted). The heptenoic acids were esterified with CH<sub>2</sub>N<sub>2</sub> (yields for the above two steps: m- 52-84 %; p- 41-72 %). Deprotection of TBS group (n-Bu<sub>4</sub>N+F-, THF, 38-100 %) and one pot formation of azide 19 via a mesylate yielded in 71-91 % the desired azides 19 and 20. Typically the (E)- and (Z)isomers were separated at this stage. The nickel chloride catalyzed NaBH<sub>4</sub> reduction of the azide to amine (47-84 %), then sulfonamide formation (39–82 %), followed by hydrolysis of the ester (44-99 %) furnished the final products 21 and 22.

Scheme II. a) n-BuLi, Et<sub>2</sub>O, -78 °C; nitrobenzaldehyde; b) MnO<sub>2</sub>, THF, reflux; c) Br Ph<sub>3</sub>P<sup>+</sup>(CH<sub>2</sub>)<sub>n+1</sub>CO<sub>2</sub>H, t-BuOK, THF, 0 °C; d) CH<sub>2</sub>N<sub>2</sub>, Et<sub>2</sub>O-THF; chromatographic separation; e) NiCl<sub>2</sub>·6 H<sub>2</sub>O, NaBH<sub>4</sub>, MeOH, 0 °C; f) p-X-PhSO<sub>2</sub>Cl, pyr.; g) 1 N NaOH, THF-MeOH (1:1); H<sup>+</sup>.

<sup>&</sup>lt;sup>b</sup>The thromboxane receptor antagonism (TRA) and thromboxane synthase inhibition (TSI) were determined in triplicate using a human platelet binding assay and human serum levels of TXB<sub>2</sub>.

Stereochemical assignments of the trisubstituted  $\omega$ -alkenoic acids rested on their  $^1H$  NMR spectra, NOE experiments (NOESY) on their key intermediates, and their mobility on silica gel. The chemical shift of the olefinic proton of E-isomers was shielded upfield compared to that of Z-isomers. The nuclear Overhauser effect (NOE) was observed between the olefinic proton of the alkenoic acid chain and the  $C_2$  and  $C_4$  protons of pyridyl ring as well as between the  $C_5$  allylic protons of the alkenoic acid chain and the  $C_2$  and  $C_6$  protons of phenyl ring for the (E)-isomer. For the (Z)-isomer, a NOE was observed between the allylic protons of alkenoic acid chain and the  $C_2$  and  $C_4$  protons of pyridyl ring, whereas NOE between the olefinic proton and the  $C_2$  and  $C_6$  protons of phenyl ring was not observable.

Mobility of the geometrical isomers indicated that, in general, the (E)-isomers were more polar on silica gel than the (Z)-isomers. Observation of <sup>1</sup>H NMR spectra and mobility of our compounds was consistent with the literature precedence. <sup>15</sup>

#### Pharmacological Results and Discussion

The series of compounds synthesized and depicted by the structure I were evaluated for their *in vitro* potency in a receptor binding assay and inhibition of  $TXA_2$  biosynthesis. Tables 1–3 list the  $K_d$  and  $IC_{50}$  values for *in vitro* TRA and TSI assays, respectively. Our initial structure and activity relationship study (SAR) in the series 9 and 10 indicated that alkenoic acid chain length had a profound effect on TSI and somewhat modest effect on TRA activity (Table 1). In the *meta* substituted series,

compounds 9c, 9d, and 9f (where n = 4, 5, and 7) were the best TRAs and TSIs (TRAs:  $K_d = 2.8-2.7 \mu M$ ; TSIs: IC<sub>50</sub> = 230–93 nM), whereas in the para series 10c (n = 5) had distinctively superior dual activities than other compounds with either shorter or longer chain length. Moreover, the octenoic acid 10c showed fairly balanced TRA and TSI activity ( $K_d = 324 \text{ nM}$ , IC<sub>50</sub> = 130 nM). The next step in our SAR was to examine the effects of substitution on the benzenesulfonamide ring and double bond geometry to improve and balance the combined activity. We chose 9c in the meta series as the base structure for this study since the distance (~9 Å) between the pyridyl nitrogen and the carboxylic acid group of the side chain has been estimated to be optimal for TSI activity. 15,20 A profound substituent effect was seen with TRA activity (Table 2). The electron withdrawing groups (with the exception of fluoro compound) appear to be more beneficial than the electron donating groups for the TRA activity. For example, the halogenated benzenesulfonamide derivatives 15e, 15f, and 15g showed far better TRA activity ( $K_d = 0.80-0.59 \mu M$ ) than others. However, the substituent effect on TSI was not as clear. Among the halogenated compounds, the TRA activity increased dramatically from the fluoro- (15d) to the iodo-substituted compound (15g), whereas the order of TSI activity was completely reversed. The double bond geometry of the alkenoic acid side chain affected both the TRA and TSI activity but more profoundly the TSI. Kato et al. have reported 15 that (E)- $\omega$ -pyridylalkenoic acids were 2-10 times better TSI than the (Z)-isomers. This appeared to be true with our compounds. The base structure 9c was prepared according to Scheme II to separate the (E/Z)-mixture. The in vitro assay showed that (E)-9c was a better TRA/TSI than (Z)-9c (Table 2). With these results, we speculated that the iodo-substituted (the best TRA) (E)-alkenoic acid (better TSI) should be a superior dual-acting agent than others in the series. This was indeed the case. The compound (E)-15g was the best TRA/TSI and showed a well-balanced TRA/TSI activity  $(K_d = 322 \text{ nM}, IC_{50} = 300 \text{ nM})$ . Notice that (E)-15b was tenfold more potent than (Z)-15b as a TSI. In the para series, we modified 10c by incorporating iodine in the benzenesulfonamide ring (16b). This resulted in a twofold improvement in both TRA and TSI activity compared to **10c** ( $K_d = 162 \text{ nM}$ ,  $IC_{50} = 68 \text{ nM}$ ).

Scheme III. a) TBSCl, imidazole, CH<sub>2</sub>Cl<sub>2</sub>; b) MnO<sub>2</sub>, THF, reflux; c) 3-lithiopyridine, Et<sub>2</sub>O, -78 °C; d) Br<sup>-</sup>Ph<sub>3</sub>P<sup>+</sup>(CH<sub>2</sub>)<sub>n+1</sub>CO<sub>2</sub>H,  $\neq$  BuOK, THF, 0 °C; e) CH<sub>2</sub>N<sub>2</sub>, Et<sub>2</sub>O-THF; f) n- Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup>, THF; g) MeSO<sub>2</sub>Cl, Et<sub>3</sub>N, PhCH<sub>3</sub>, 0 °C; aq. NaN  $\neq$  n- Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup>, 60 °C; chromatographic separation; h) NiCl<sub>2</sub>·6 H<sub>2</sub>O, NaBH<sub>4</sub>, MeOH, 0 °C; i) p-X-PhSO<sub>2</sub>Cl, pyr.; j) 1 N NaOH, THF-MeOH (1:1); H<sup>+</sup>.

Table 2. In vitro activities of compounds 15 and 16

| Compounds    | n | x   | EIZ     | TRA<br><u>Kd (µM)</u> b | TSI<br>IC <u>50 (nM</u> )b |
|--------------|---|-----|---------|-------------------------|----------------------------|
| 9c           | 4 | н   | Z       | $5.8 \pm 0.6$           | 498 ± 180                  |
| 15a          | 4 | Me  | Z       | $2.3\pm0.2$             | 840                        |
| 1 <b>5</b> b | 4 | OMe | Z       | 5.4 ± 0.5               | 3,100                      |
| 15c          | 4 | CF3 | z       | 1.1 ± 0.2               | 470 ± 110                  |
| 15d          | 4 | F   | Z       | 3.0 ± 0.3               | 300                        |
| 15e          | 4 | Cl  | Z       | $0.80 \pm 0.06$         | 900                        |
| 15f          | 4 | Br  | z       | $0.72 \pm 0.09$         | 1,030 ± 270                |
| 15g          | 4 | I   | Z       | $0.59 \pm 0.04$         | 4,000                      |
| 15h          | 4 | Ph  | Z       | $2.01 \pm 0.35$         | 1,000 ± 50                 |
| 16a          | 4 | I   | Z       | 0.162±0.025             | 4,300                      |
| 9c           | 4 | H   | E       | $2.6\pm0.3$             | 300                        |
| 15g          | 4 | 1   | E       | $0.322 \pm 0.021$       | 300                        |
| 16b          | 5 | I   | >96%E a | 0.162±0.015             | 68 ± 21                    |

<sup>a</sup>The (E/Z) ratio was determined by HPLC analysis.

<sup>b</sup>The thromboxane receptor antagonism (TRA) and thromboxane synthase inhibition (TSI) were determined in triplicate using a human platelet binding assay and human serum levels of TXB<sub>2</sub>.

The third step of our SAR study was to give more conformational flexibility to the arylsulfonamido group. This was done by incorporating a methylene chain between the sulfonamide nitrogen and the phenyl ring attached to the alkenoic acid chain, as in the series 21 and 22 (Scheme III). Such flexibility in the molecule may optimize the fit of the inhibitor in the enzyme active site as well as at the receptor site. Our results show that this goal was apparently realized (Table 3). A dramatic improvement in both TRA and TSI was seen in meta series: for example, 9b to 21a, 9c to 21b, 9d to 21d, and 15g to 21c. The effects of changing double bond geometry and chain length of the alkenoic acids in the meta series 21 were significantly diminished, though the (E)-isomers were still preferred and n = 4 was optimal chain length. The effect of iodo-substitution for TRA resulted in about five-fold improvement (21b to 21c), significant but not as much as in the series lacking the methylene chain; i.e. 9c to 15g (8~10-fold). Compound (E)-21c was identified as the most potent TRA/TSI ( $K_d = 53 \text{ nM}$ , IC<sub>50</sub> = 23 nM) in this series. The dramatic improvement by incorporating a methylene chain was also observed in the para series except where n = 5; for example, 10a to (E)-22a and 10b to (E)-22b. Interestingly, the effect of double bond geometry of the alkenoic acid side chain for both TRA and TSI was more pronounced in the para series 22 than in the

Table 3. In vitro activities of compounds 21 and 22

| Compounds | a | x | <u>E/Z</u> a  | TRA<br><u>Kd (uM)</u> | TSI<br>IC50 (pM) |
|-----------|---|---|---------------|-----------------------|------------------|
| 21a       | 3 | H | >97.6%E       | $0.310\pm0.038$       | 32 ± 15          |
|           |   |   | >95.5%Z       | $0.265 \pm 0.040$     | 205 ± 40         |
| 21b       | 4 | H | 76:24         | $0.280 \pm 0.051$     | 30 ± 10          |
|           |   |   | 9:91          | $0.700 \pm 0.110$     | 67 ± 15          |
| 21c       | 4 | I | 86 : 14       | $0.053 \pm 0.008$     | 23 ± 4           |
|           |   |   | 21 : 79       | $0.131 \pm 0.026$     | 53 ± 9           |
| 214       | 5 | н | 50 : 50       | $0.312 \pm 0.047$     | 38 ± 18          |
|           |   |   | 94%Z          | $0.708 \pm 0.080$     | 336 ± 124        |
| 22a       | 3 | H | 91 <b>%</b> E | $0.380 \pm 0.020$     | 44 ± 7           |
|           |   |   | >99%Z         | $2.100 \pm 0.260$     | 550 ± 200        |
| 22b       | 4 | H | >81%E         | $0.410 \pm 0.050$     | 49 ± 7           |
|           |   |   | >98%Z         | $2.660 \pm 0.850$     | 290 ± 160        |
| 22c       | 4 | I | 97%Z          | $0.220 \pm 0.010$     | 390 ± 10         |
| 224       | 5 | H | ≥69%E         | $0.580 \pm 0.030$     | 28 ± 6           |
|           |   |   | >91 <b>%Z</b> | $1.260 \pm 0.090$     | 750 ± 160        |

The (E/Z) ratio was determined by HPLC analysis.

<sup>b</sup>The thromboxane receptor antagonism (TRA) and thromboxane synthase inhibition (TSI) were determined in triplicate using a human platelet binding assay and human serum levels of TXB<sub>2</sub>.

meta series 21. However, length of the alkenoic chain had little effect and the little effect observed was inconsistent in the para series 22. The beneficial effect of iodosubstitution for TRA was observed in this series also ((Z)-22b to 22c). Of these homologated series, the (E)-isomers were more potent than the (Z)-isomers and the meta series 21 was more potent than the para series 22 for both TRA and TSI activity in all cases. The iodo-substituent on the benzenesulfonamide, while affecting the TRA significantly, did not influence the TSI activity appreciably even in the case of (Z)-isomer (21b vs 21c, (Z)-22b vs 22c)in these homologated series. It is possible that when the pyridine ring and the carboxylic acid group bind to the enzyme, 15 the sulfonamide chain with its conformational flexibility orientates itself in such a way that both electronic and steric variations at the end of its chain have minimal effect on the enzyme inhibitory activity. Neither the series 9 and 10 where the changes in the alkenoic acid side chain length became an important factor, nor the series 15 and 16 where the (E)-geometry of the double bond became significant, had this flexibility. Indeed, such inflexibility in these compounds most likely limited their TRA/TSI activity.

One of the advantages of TSI agents is the so-called 'shunt' effect whereby accumulated  $PGH_2$ , resulting from inhibition of  $TXA_2$  biosynthesis by the agent, is redirected to  $PGI_2$  synthesis. We have examined whether such an effect is indeed manifested by our dual-acting compounds. The result is shown in Figure 1. The compound 9 c significantly increased the production of serum  $PGI_2$  in vitro as measured by the amount of 6-keto- $PGF_{1\alpha}$ , a stable metabolite of  $PGI_2$ , produced. In contrast, aspirin abolished  $PGI_2$  formation via its action against cyclooxygenase.

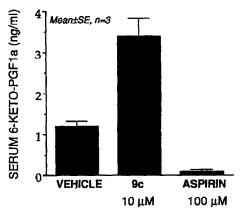


Figure 1. Effect of 9c and aspirin on human serum prostacyclin levels.

Another important aspect in the development of TRA/TSI agents is that the compound should be orally active. We felt that the oral activity of the agent ought to be examined in the early stage of drug development. Therefore the ex vivo activity of compounds 9c and 10c via the oral route was determined in rats. ED<sub>50</sub>s for TSI after 1 h dosing were ~1.0 mg/kg for 9c and ~1.2 mg/kg for 10c.

#### Conclusion

The series of compounds described in this paper, designed by 'hybridizing' two prototypical representatives of the TRA and TSI pharmacophores were found to exhibit dual TRA/TSI activities. Changes in the chain length as well as the trisubstituted olefin geometry of ω-pyridylalkenoic acid side chain had significant effects on both TXA2 biosynthesis<sup>21</sup> and TXA<sub>2</sub> receptor blockade. The substitution pattern of the arylsulfonamido group profoundly affected the TXA<sub>2</sub> receptor blockade.<sup>22</sup> Among the various substituents, iodo-substitution gave the most potent compound. The flexibility between the arylsulfonamido group and the phenyl ring attached to the alkenoic acid side chain significantly enhanced both activities. The compound (E)-21c was identified as the most potent TRA/TSI ( $K_d = 53 \text{ nM}$ , IC<sub>50</sub> = 23 nM) in the series studied. The compounds 9c and 10c have indicated that members of this series of compounds also exhibit the so-called 'shunt' effect by showing a lack of cyclooxygenase inhibition while promoting PGI<sub>2</sub> production in vitro. In addition, they have demonstrated activity following oral administration.

These results indicate that type I compounds have therapeutic potential as orally active, dual-acting TRA/TSI agents for cardiovascular and renal diseases. <sup>23</sup>

# **Experimental Section**

# General procedure

All solvents and reagents were purchased from commercial sources and used as received, unless otherwise indicated. Tetrahydrofuran (THF) was distilled from sodium benzophenone ketyl prior to use. Triethylamine and pyridine were distilled from CaH<sub>2</sub> and stored over 4 Å molecular sieves. Benzene and toluene were dried over sodium metal. All reactions were performed under a positive pressure of dry nitrogen. The 'preparative HPLC' was performed on a Waters PrepLC System 500A with the solvent indicated. Analytical HPLC was carried out on a Waters Model 510 using Nova C<sub>18</sub> column with CH<sub>3</sub>CN-MeOH-H<sub>2</sub>O solvent system which contained 0.5 % NH<sub>4</sub>OAc or Chiralcel OD-R column with CH<sub>3</sub>CN-H<sub>2</sub>O solvent system which contained 0.1 % NaClO<sub>4</sub>. Flash chromatography was carried out on E. Merck Kieselgel 60 (230-400 mesh). <sup>1</sup>H NMR spectra were recorded on a GE Q(E)-300 (routine) and on Bruker AM-500 (NOE) spectrometer. The chemical shifts are given in  $\delta$  values relative to residual proton resonances of the deuterated solvents used (CDCl<sub>3</sub> 7.26, DMSO-d<sub>6</sub> 2.49, acetone-d<sub>6</sub> 2.05). Analysis of the (E/Z) isomeric ratio of alkenoic acids was accomplished either by analytical HPLC or <sup>1</sup>H NMR. In the latter method, the ratio was obtained from integration of the olefinic proton of the product acids or their esters. The chemical purity of isomeric product mixtures was determined by analytical HPLC where the hygroscopic nature of these compounds made correct elemental analysis difficult. Field desorption (FDMS) and fast atom bombardment mass spectra (FABMS) were obtained on a VG ZAB-3F or VG 70-SE instrument.

# 3-[(Benzenesulfonyl)amino]phenyl 3-pyridyl ketone (7)

To a solution of 2.5 g (8.0 mmol) of 3-[(benzenesulfonyl)amino]bromobenzene in 80 mL of THF was added dropwise 10.0 mL (16.0 mmol) of 1.6 M n-BuLi at -78 °C. After stirring for 75 min, 1.8 mL (19.2 mmol) of 3-pyridinecarboxaldehyde was added and the mixture was stirred at -78 °C for 2.5 h. The reaction was quenched with 80 mL of saturated aqueous NH<sub>4</sub>Cl and extracted with  $3 \times 110$  mL of EtOAc. The extract was dried over MgSO<sub>4</sub>, concentrated, and purified by flash chromatography using 3 % MeOH–CH<sub>2</sub>Cl<sub>2</sub> to yield 1.62 g (60 %) of 3-(benzenesulfonylamino)phenyl(3-pyridyl)carbinol. The carbinol (1.60 g) was oxidized by Swern's method<sup>24</sup> to furnish 0.67 g (42 %) of the desired ketone 7: <sup>1</sup>H NMR (DMSO)  $\delta$  10.55 (s, 1H), 8.79 (br d, J = 4.3 Hz, 1H), 8.73 (s, 1H), 7.93 (d, J = 8.0 Hz, 1H), 7.72 (d, J = 7.8Hz, 2H), 7.60-7.39 (m, 8H); FDMS 338 (M<sup>+</sup>).

#### 4-[(Benzenesulfonyl)amino]phenyl 3-pyridyl ketone (8)

4-[(Benzenesulfonyl)amino]phenyl(3-pyridyl)carbinol was prepared as above from 2.5 g (8.0 mmol) of 4-[(benzenesulfonyl)amino]bromobenzene in 59 % yield. The carbinol, 1.36 g (4.0 mmol), was oxidized with 5.44 g (4 × by weight) of MnO<sub>2</sub> in 50 mL of 1,4-dioxane at 70 °C (bath temperature) for 9 h. The removal of the oxidant by filtration through a silica gel-Celite pad and of the solvent under reduced pressure yielded 1.30 g (96 %) of the clean crude ketone 8 which was used without further purification:  $^1$ H NMR (DMSO)  $\delta$  10.96 (s, 1H), 8.75 (br s, 1H), 8.74 (s, 1H), 7.98 (dt, J = 7.9, 1.8 Hz, 1H), 7.81 (d, J = 6.9 Hz, 2H), 7.65 (d, J = 8.6 Hz, 2H), 7.56 (m, 4H), 7.24 (d, J = 8.6 Hz, 2H); FDMS 338 (M<sup>+</sup>). Anal. (C<sub>18</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub>S) C, H, N.

#### General procedure for Wittig reactions (A)

5-[3-[(Benzenesulfonyl)amino]phenyl]-5-(3-pyridyl)-pent-4-enoic acid (9a). A suspension of 2.93 g (6.82 mmol) of (3-carboxypropyl)triphenylphosphonium bromide in 7.5 mL of THF was treated with 13.7 mL (13.7 mmol) of 1.0 M sodium hexamethyldisilazide (NaHMDS) in THF at -78 °C for 2 h 45 min. To this ylide solution was then added 1.0 g (3.10 mmol) of 3-[(benzenesulfonyl)amino]phenyl 3pyridyl ketone (7) in 8 mL of THF and the mixture was stirred at -78 °C for 22 h, warmed to room temperature and stirred for an additional 2 h. The reaction was then quenched with 30 mL of brine. The mixture was concentrated to dryness, suspended in MeOH, and filtered. The filtrate was concentrated and purified by flash chromatography using MeOH-Acetone-CH<sub>2</sub>Cl<sub>2</sub> (5:5:90) to give 0.605 g (48 %) of the Wittig product 9a in that (E)isomer predominated: FDMS 409 (M+1). (C<sub>22</sub>H<sub>20</sub>N<sub>2</sub>O<sub>4</sub>S) C, H, N: calcd, 6.86; found, 5.03. 98.5 % Pure by analytical HPLC.

6-[3-[(Benzenesulfonyl) amino]p heyl]-6-(3-pyridyl)-hex-5-enoic acid (9b). Prepared as above from 975.3 mg (2.2 mmol) of (4-carboxybutyl)triphenylphosphonium bromide, 4.4 mL (4.4 mmol) of 1.0 M NaHMDS in THF and 340 mg (1.0 mmol) of the ketone 7 in 55 % yield: FDMS 423 (M+1). 97.6 % Pure by analytical HPLC.

7-[3-[(Benzenesulfonyl)amino]phenyl]-7-(3-pyridyl)-hept-7-enoic acid (9c). Prepared as above from 3.41 g (7.46 mmol) of (5-carboxypentyl)triphenylphosphonium bromide, 15.0 mL (15.0 mmol) of 1.0 M NaHMDS in THF and 1.14 g (3.37 mmol) of the ketone 7 in 54 % yield: FDMS 437 (M+1). Anal. ( $C_{24}H_{24}N_{2}O_{4}S$ ) C, H, N. 100 % Pure by analytical HPLC.

8-[3-[(Benzenesulfonyl)amino]phenyl] -8-(3-pyridyl)-oct-7-enoic acid (9d). Prepared as above from 623 mg (1.32 mmol) of (6-carboxyhexyl)triphenylphosphonium bromide, 2.64 mL (2.64 mmol) of 1.0 M NaHMDS in THF and 203 mg (0.6 mmol) of the ketone 7 in 9 % yield: FDMS 451 (M+1). 99.0 % Pure by analytical HPLC.

9-[3-[(Benzenesulfonyl)amino]phenyl]-9-(3-pyridyl)-non-8-enoic acid (9e). Prepared as above from 880 mg (1.8

mmol) of (7-carboxyheptyl)triphenylphosphonium bromide, 3.6 mL (3.6 mmol) of 1.0 M NaHMDS in THF and 205 mg (0.6 mmol) of the ketone 7 in 34 % yield: FDMS 465 (M+1). 97.4 % Pure by analytical HPLC.

10-[3-[(Benzenesulfonyl) amino] pheny]-10-(3-pyridyl)-dec-9-enoic acid (9f). Prepared as above from 2.25 g (3.75 mmol) of [9-(t-butyldimethylsiloxy))nonyl]triphenyl-phosphonium bromide, 7.5 mL (7.5 mmol) of 1.0 M NaHMDS in THF and 573 mg (1.7 mmol) of the ketone 7 to yield 10-[3-[(benzenesulfonyl)amino]phenyl]-10-(3-pyridyl)-dec-9-enyl t-butyldimethylsilyl ether which was oxidized with Jones reagent (2 equiv.) in acetone at 0 °C for 2 h and purified by flash chromatography using EtOH-i-PrOH-CH<sub>2</sub>Cl<sub>2</sub> (5:5:90) to provide the decenoic acid 9f (2%): FDMS 479 (M+1). 91.0 % Pure by analytical HPLC.

General procedure for Wittig reactions (B).

6-[4-[(Benzenesulfonyl)amino]phenyl]-6-(3-pyridyl)-hex-5-enoic acid (10a). To a suspension of 203 mg (0.6 mmol) of 4-[(benzenesulfonyl)amino]phenyl 3-pyridyl ketone (8) and 586 mg (1.32 mmol) of (4-carboxybutyl)triphenylphosphonium bromide in 3.0 mL of THF was added dropwise 2.6 mL (2.60 mmol) of 1.0 M t-BuOK in THF at -10 °C. The mixture was stirred at -9 to 14 °C overnight and then allowed to warm to room temperature and stirred for another 20 h. The reaction mixture was then quenched with 1.0 N HCl while adjusting pH to ~7, concentrated, and purified by flash chromatography using MeOH-AcOH-CH<sub>2</sub>Cl<sub>2</sub> (5:0.5:94.5) to afford 233.3 mg (92 %) of the hexenoic acid 10a:  ${}^{1}$ H NMR (CDCl<sub>2</sub>)  $\delta$  8.42 (br d, J = 4.5 Hz, 1H), 8.39 (br s, 1H), 7.78 (br d, J = 7.5Hz, 2H), 7.44 (m, 5H), 7.20 (dd, J = 7.9, 5.0 Hz, 1H), 7.10 (d, J = 8.4 Hz, 2H), 6.96 (d, J = 8.3 Hz, 2H), 6.06 (t, J = 8.4 Hz, 2H)7.4 Hz, 1H), 2.30 (dd, J = 7.4, 7.3 Hz, 2H), 2.12 (ddd, J =7.5, 7.3, 7.2 Hz, 2H), 1.70 (m, 2H); FDM\$ 423 (M+1). Anal.  $(C_{23}H_{22}N_2O_4S)$  C, H, N: calcd, 6.63; found, 5.25. 98.4 % Pure by analytical HPLC.

7-[4-[(Benzenesulfonyl)amino]phenyl]-7-(3-pyridyl)-hept-6-enoic acid (10b). Prepared as above from 205 mg (0.61 mmol) of the ketone 8, 605 mg (1.32 mmol) of (5-carboxypentyl)triphenylphosphonium bromide, and 2.65 mL (2.65 mmol) of 1.0 M t-BuOK in THF in 88 % yield: FDMS 437 (M+1). 91.9 % Pure by analytical HPLC.

8-[4-[(Benzenesulfonyl)amino]phenyl] -8-(3-pyridyl)-oct-7-enoic acid (10c). Prepared as above from 205 mg (0.61 mmol) of the ketone 8, 627 mg (1.33 mmol) of (6-carboxyhexyl)triphenylphosphonium bromide, and 2.65 mL (2.65 mmol) of 1.0 M t-BuOK in THF in quantitative yield: FDMS 451 (M+1). 98.0 % Pure by analytical HPLC.

9-[4-[(Benzenesulfonyl)amino]phenyl]-9-(3-pyridyl)-non-8-enoic acid (10d). Prepared as above from 340 mg (1.0 mmol) of the ketone 8, 1.46 g (3.0 mmol) of (7-carboxyheptyl)triphenylphosphonium bromide, and 6.0 mL (6.0 mmol) of 1.0 M t-BuOK in THF in quantitative yield: FDMS 465 (M+1). 95.3 % Pure by analytical HPLC.

# 3-Nitrophenyl 3-pyridyl ketone (11)

To a solution of 5.13 g (32.5 mmol) of 3-bromopyridine in 100 mL of anhydrous Et<sub>2</sub>O was added dropwise 20.0 mL (32.5 mmol) of 1.6 M n-BuLi in hexanes at -78 °C over a 20 min period. After stirring for 30 min, 4.91 g (32.5 mmol) of 3-nitrobenzaldehyde in 90 mL of anhydrous Et<sub>2</sub>O was cannulated into the yellow slurry solution. The mixture was stirred at -78 °C for 2.5 h. The cold bath was removed and the reaction was quenched with 100 mL of brine after 10 min. The mixture was extracted with 250-500 mL of  $CH_2Cl_2$  (4 ×) which was washed once with 100 mL of brine. The extracts were dried over MgSO<sub>4</sub>. concentrated and purified by preparative HPLC with 50 % EtOAc-CH<sub>2</sub>Cl<sub>2</sub> to afford 4.05 g (54 %) of the 3nitrophenyl(3-pyridyl)carbinol. The carbinol was oxidized by Swern's method to the ketone 11 in 98 % yield: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.98 (s, 1H), 8.86 (d, J = 4.0 Hz, 1H), 8.62 (dd, J = 1.8, 1.5 Hz, 1H), 8.48 (m, 1H), 8.13 (m, 2H), 7.74 (t, J = 7.9 Hz, 1H), 7.50 (dd, J = 7.8, 4.8 Hz, 1H).

# 4-Nitrophenyl 3-pyridyl ketone (12)

4-Nitrophenyl(3-pyridyl)carbinol was prepared similarly to the above from 5.59 g (35.4 mmol) of 3-bromopyridine treated with 24.5 mL (39.2 mmol) of 1.6 M n-BuLi and 5.88 g (38.9 mmol) of 4-nitrobenzaldehyde in 35 % yield. The carbinol, 2.86 g (12.4 mmol) was oxidized with 11.44 g of MnO<sub>2</sub> in 100 mL of THF at reflux for 2 h. Removal of the oxidant by filtration through a silica gel-Celite pad and of the solvent under reduced pressure yielded 2.70 g (95 %) of the clean crude ketone 12 which was used without further purification:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.99 (d, J = 1.8 Hz, 1H), 8.87 (dd, J = 4.6, 1.4 Hz, 1H), 8.38 (d, J = 8.6 Hz, 2H), 8.15 (dt, J = 7.9, 1.8 Hz, 1H), 7.98 (d, J = 8.6 Hz, 2H), 7.52 (m, 1H).

Methyl (E)- and (Z)-7-(3-nitrophenyl)-7-(3-pyridyl)-hept-6-enoates (13c)

7-(3-Nitrophenyl)-7-(3-pyridyl)-hept-6-enoic acid was prepared from 1.1544 g (5.06 mmol) of 3-nitrophenyl 3pyridyl ketone (11), 4.6274 g (10.1 mmol) of (5carboxypentyl)triphenylphosphonium bromide and 10.5 mL (10.5 mmol) of 1.0 M t-BuOK in THF according to the Wittig reaction method (B). Flash chromatography with MeOH-Et<sub>2</sub>O-CH<sub>2</sub>Cl<sub>2</sub> (5:15:80) gave ca 880 mg of the acid which was esterified with CH<sub>2</sub>N<sub>2</sub> by a standard method and purified by flash chromatography with Et<sub>2</sub>O-Hex-CH<sub>2</sub>Cl<sub>2</sub> (1:1:1) to yield 601.3 mg of the (Z)heptenoate (less polar material) and 206.5 mg of (E)heptenoate (45 % in total for two steps). (E)-isomer: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.46 (m, 2H), 8.19 (dd, J = 8.0, 1.3 Hz, 1H), 8.02 (br s, 1H), 7.57 (dd, J = 8.0, 7.8 Hz, 1H), 7.48 (br d, J = 7.3 Hz, 1H), 7.41 (dt, J = 7.9, 1.9 Hz, 1H), 7.19 (dd, J = 5.0, 3.0 Hz, 1H), 6.19 (t, J = 7.5 Hz, 1H), 3.63 (s, 3H), 2.26 (dd, J = 7.3, 7.1 Hz, 2H), 2.14 (ddd, J = 7.4, 7.3, 7.3 Hz, 2H), 1.64 (m, 2H), 1.49 (m, 2H); FDMS 340 (M<sup>+</sup>). (Z)-isomer: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.59 (dd, J = 4.9, 1.3 Hz, 1H), 8.44 (br d, J = 1.7 Hz, 1H), 8.06 (m, 2H), 7.43 (m, 3H), 7.34 (dd, J = 7.9, 4.6 Hz, 1H), 6.27 (t, J = 7.5 Hz,

1H), 3.64 (s, 3H), 2.26 (dd, J = 7.0, 6.4 Hz, 2H), 2.15 (ddd, J = 7.4, 7.3, 7.3 Hz, 2H), 1.62 (m, 2H), 1.51 (m, 2H); FDMS 340 (M<sup>+</sup>).

Methyl (E)- and (Z)-7-(4-nitrophenyl)-7-(3-pyridyl)-hept-6-enoates (14b)

7-(4-Nitrophenyl)-7-(3-pyridyl)-hept-6-enoic acid was prepared similarly to the above from 518.2 mg (2.27 mmol) of 4-nitrophenyl 3-pyridyl ketone (12), 2.077 g (4.54 mmol) of (5-carboxypentyl)triphenylphosphonium bromide and 4.5 mL (4.5 mmol) of 1.0 M t-BuOK in THF. The esterification of the (E/Z)-mixture of acids yielded 368.4 mg of (Z)-heptenoate and 69.3 mg of (E)-heptenoate (57 % in total for two steps). (E)-isomer: <sup>1</sup>H NMR  $(CDCl_3) \delta$  8.46 (br s, 2H), 8.24 (d, J = 8.6 Hz, 2H), 7.39 (br d, J = 8.0 Hz, 1H), 7.33 (d, J = 8.6 Hz, 2H), 7.20 (m, 1H), 6.19 (t, J = 7.5 Hz, 1H), 3.64 (s, 3H), 2.26 (dd, J =7.4, 7.1 Hz, 2H), 2.14 (ddd, J = 7.4, 7.3, 7.3 Hz, 2H), 1.60 (m, 2H), 1.50 (m, 2H). (Z)-isomer:  ${}^{1}H$  NMR (CDCl<sub>3</sub>)  $\delta$ 8.59 (br d, J = 1.9 Hz, 1H), 8.42 (br s, 1H), 8.10 (d, J = 8.7Hz, 2H), 7.44 (br dd, J = 7.8, 1.5 Hz, 1H), 7.34 (m, 1H), 7.30 (d, J = 8.7 Hz, 2H), 6.32 (t, J = 7.5 Hz, 1H), 3.62 (s, 3H), 2.25 (dd, J = 7.4, 7.0 Hz, 2H), 2.14 (ddd, J = 7.4, 7.3, 7.2 Hz, 2H), 1.59 (m, 2H), 1.49 (m, 2H).

#### General procedure for the alkenoic acids

(Z)-7-[3-[4-(Toluenesulfonyl)amino]phenyl]-7-(3-pyridyl)hept-6-enoic acid (15a). To a pale green, clear solution of 525.1 mg (1.54 mmol) of methyl (Z)-7-(3-nitrophenyl)-7-(3-pyridyl)-hept-6-enoate ((Z)-13c) and 366.5 mg (1.54)mmol) of NiCl<sub>2</sub>·6H<sub>2</sub>O in 14 mL of MeOH at 0 °C was added 292 mg (7.11 mmol) of NaBH<sub>4</sub> in small portions over a 30 min period. After stirring for 30 min, the solvent was removed under reduced pressure and the residue was treated with ca 5 mL of 5 N HCl and then basified with ca 30 mL of 28 % NH<sub>4</sub>OH at 0 °C. The clear purple solution was then extracted with 3 × 50 mL of CH<sub>2</sub>Cl<sub>2</sub> which was washed with ca 30 mL of brine. The extract was dried over MgSO<sub>4</sub>, concentrated, and purified by flash chromatography using 75 % EtOAc-hexanes to afford 420.5 mg (88 %) of methyl (Z)-7-(3-aminophenyl)-7-(3-pyridyl)-hept-6-enoate: <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 8.53 (dd, J = 4.9, 1.5 Hz, 1H), 8.43 (d, J = 1.8 Hz, 1H), 7.45 (dt. J =7.8, 1.7 Hz, 1H), 7.27 (m, 1H), 7.04 (t, J = 7.8 Hz, 1H), 6.56 (m, 2H), 6.45 (dd, J = 1.9, 1.7 Hz, 1H), 6.11 (t, J =7.5 Hz, 1H), 3.83 (s, 3H), 3.61 (br s, 2H), 2.25 (dd, J = 7.5, 7.2 Hz, 2H), 2.08 (ddd, J = 7.4, 7.3, 7.3 Hz, 2H), 1.60 (m, 2H), 1.46 (m, 2H); FDMS 310 (M+).

A solution of 34.4 mg (0.11 mmol) of the above amine and 42.3 mg (0.22 mmol) of 4-toluenesulfonyl chloride in 0.5 mL of pyridine was stirred at 0–10 °C for 4 h. The mixture was concentrated *in vacuo* and the residue was purified by flash chromatography with 30 % EtOAc–CH<sub>2</sub>Cl<sub>2</sub> to afford 45.5 mg (0.098 mmol) of the corresponding sulfonamide ester which was dissolved in 0.5 mL of THF and hydrolyzed with 200  $\mu$ L of 1.0 N NaOH (0.20 mmol) at room temperature overnight. After the solution was neutralized with 200  $\mu$ L of 1.0 N HCl at 0

°C and concentrated to dryness, the crude product was suspended in  $CH_2Cl_2$  and filtered through a filter paper to remove the NaCl salt formed. The filtrate was concentrated in vacuo to yield 41.8 mg (84 % for 2 steps) of the desired acid 15a:  $^1H$  NMR (CDCl<sub>3</sub>)  $\delta \sim 10.1$  (very br s, 1H), 8.51 (br s, 1H), 8.33 (br s, 1H), 7.58 (d, J=7.9 Hz, 2H), 7.44 (distorted br d, J=7.3 Hz, 1H), 7.31 (br s, 1H), 7.15 (d, J=7.9 Hz, 2H), 7.07 (br dd, J=7.9, 7.9 Hz, 1H), 6.99 (distorted br d, J=7.7 Hz, 1H), 6.83 (br s, 2H), 6.06 (t, J=7.3 Hz, 1H), 2.31 (s, 3H), 2.27 (buried, 2H), 2.03 (m, 2H), 1.59–1.45 (m, 4H); FDMS 451 (M+1). Anal. ( $C_{25}H_{26}N_2O_4S$ ) C, H, N.

(Z)-7-[3-[4-(Methoxybenzenesulfonyl)amino]phenyl]-7-(3-pyridyl)-hept-6-enoic acid (15b). Prepared as above in 3 steps from (Z)-13c and 4-methoxybenzenesulfonyl chloride (79 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  ~9.50 (very br s, 1H), 8.50 (br s, 1H), 8.34 (br s, 1H), 7.63 (d, J = 8.9 Hz, 2H), 7.43 (distorted d, J = 7.7 Hz, 1H), 7.33 (br s, 1H), 7.08 (dd, J = 7.8, 7.7 Hz, 1H), 7.00 (distorted d, J = 8.1 Hz, 1H), 6.83 (d, J = 8.7 Hz, 2H), 6.83 (buried, 2H), 6.07 (t, J = 7.4 Hz, 1H), 3.77 (s, 3H), 2.28 (dd, J = 7.1, 6.9 Hz, 2H), 2.05 (ddd, J = 7.2, 7.2, 6.9 Hz, 2H), 1.62–1.41 (m, 4H); FDMS 466 (M<sup>+</sup>). Anal. (C<sub>25</sub>H<sub>26</sub>N<sub>2</sub>O<sub>5</sub>S) C, H, N.

(Z) -7- [3- [[4- (Trifluoromethyl) benzenesulfonyl] amino]-phenyl]-7-(3-pyridyl)-hept-6-enoic acid (15c). Prepared as above in 3 steps from (Z)-13c and 4-(trifluoromethyl)benzenesulfonyl chloride (80 %):  $^{1}H$  NMR (CDCl<sub>3</sub>)  $\delta$  8.49 (br d, J = 1.3 Hz, 1H), 8.41 (buried, 1H), 8.33 (br s, 1H), 7.83 (d, J = 8.2 Hz, 2H), 7.62 (d, J = 8.3 Hz, 2H), 7.42 (br d, J = 7.6 Hz, 2H), 7.31 (m, 1H), 7.09 (m, 2H), 6.89 (br d, J = 7.3 Hz, 1H), 6.79 (s, 1H), 6.09 (t, J = 7.3 Hz, 1H), 2.24 (br t, 2H), 2.02 (br d, 2H), 1.58–1.42 (m, 4H); FDMS 505 (M+1).

(Z)-7- [3-[(4-Fluorobenzenesulfonyl)amino] phenyl]-7-(3-pyridyl)-hept-6-enoic acid (15d). Prepared as above in 3 steps from (Z)-13c and 4-fluorobenzenesulfonyl chloride (73%):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.49 (br s, 1H), 8.31 (s, 1H), 7.69 (dd, J = 8.7, 5.0 Hz, 2H), 7.43 (br d, J = 7.7 Hz, 1H), 7.31 (m, 1H), 7.13–6.99 (m, 4H), 6.88 (d, J = 7.6 Hz, 1H), 6.77 (s, 1H), 6.07 (t, J = 7.4 Hz, 1H), 2.27 (distorted dd, J = 6.7, 6.5 Hz, 2H), 2.04 (distorted ddd, J = 7.2, 7.1, 6.8 Hz, 2H), 1.61–1.41 (m, 2H); FDMS 455 (M<sup>+</sup>). Anal. (C<sub>24</sub>H<sub>23</sub>FN<sub>2</sub>O<sub>4</sub>S) C, H, N.

(Z)-7-[3-[(4-Chlorobenzenesulfonyl)amino]phenyl]-7- (3-pyridyl)-hept-6-enoic acid (15e). Prepared as above in 3 steps from (Z)-13c and 4-chlorobenzenesulfonyl chloride (61 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  ~9.5 (very br s, 1H), 8.49 (br d, J = 3.4 Hz, 1H), 8.32 (s, 1H), 7.61 (d, J = 8.5 Hz, 2H), 7.42 (br d, J = 7.7 Hz, 1H), 7.32 (d, J = 8.5 Hz, 2H), 7.32 (buried, 1H), 7.11 (dd, J = 7.8, 7.8 Hz, 1H), 7.02 (d, J = 8.0 Hz, 1H), 6.89 (d, J = 7.6 Hz, 1H), 6.76 (s, 1H), 6.07 (t, J = 7.4 Hz, 1H), 2.27 (dd, J = 6.8, 6.4 Hz, 2H), 2.04 (ddd, J = 7.1, 6.9, 6.8 Hz, 2H), 1.61-1.41 (m, 4H); FDMS 471 (M+1). Anal. (C<sub>24</sub>H<sub>23</sub>ClN<sub>2</sub>O<sub>4</sub>S) C, H, N.

(Z)-7-[3-[(4-Bromobenzenesulfonyl) amino] phenyl] -7-(3-pyridyl)-hept-6-enoic acid (15f). Prepared as above in 3 steps from (Z)-13c and 4-bromobenzenesulfonyl chloride

(59 %):  ${}^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.52 (br d, J = 3.5 Hz, 1H), 8.34 (s, 1H), 7.55 (d, J = 8.8 Hz, 2H), 7.50 (d, J = 8.7 Hz, 2H), 7.42 (d, J = 7.8 Hz, 1H), 7.33 (m, 1H), 7.13 (dd, J = 7.9, 7.8 Hz, 1H), 7.04 (d, J = 8.4 Hz, 1H), 6.90 (d, J = 7.7 Hz, 1H), 6.76 (s, 1H), 6.07 (t, J = 7.5 Hz, 1H), 2.27 (dd, J = 7.0, 6.9 Hz, 2H), 2.05 (br q, J = ~7.1 Hz, 2H), 1.61–1.43 (m, 4H); FDMS 515 (M $^{+}$ ).

(Z)-7- [3- [(4-Iodobenzenesulfonyl) amino] phenyl] -7- (3-pyridyl)-hept-6-enoic acid (15g). Prepared as above in 3 steps from (Z)-13c and 4-iodobenzenesulfonyl chloride (50%):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.60 (very br s, 1H), 8.53 (br d, J = 1.9 Hz, 1H), 8.36 (s, 1H), 7.73 (d, J = 8.4 Hz, 2H), 7.46-7.34 (m, 3H), 7.40 (d, J = 8.4 Hz, 2H), 7.15 (dd, J = 7.9, 7.8 Hz, 1H), 7.06 (distorted d, J = 8.3 Hz, 1H), 6.92 (d, J = 7.6 Hz, 1H), 6.78 (s, 1H), 6.10 (t, J = 7.5 Hz, 1H), 2.32 (distorted dd, J = 6.9, 6.5 Hz, 2H), 2.08 (distorted ddd, J = 7.2, 7.1, 7.0 Hz, 2H), 1.65-1.44 (m, 4H); FDMS 563 (M+1). Anal. ( $C_{24}H_{23}IN_2O_4S$ ) H, N, C: calcd, 51.25; found, 52.05.

(Z) -7- [3- [(4-Biphenylylsulfonyl) amino] phenyl] -7- (3-pyridyl)-hept-6-enoic acid (15h). Prepared as above in 3 steps from (Z)-13c and 4-biphenylylsulfonyl chloride (61%):  $^1$ H NMR (CDCl<sub>3</sub>)  $\delta$  ~9.45 (very br s, 1H), 8.47 (d, J = 4.1 Hz, 1H), 8.34 (s, 1H), 7.76 (d, J = 8.4 Hz, 2H), 7.57 (d, J = 8.4 Hz, 2H), 7.52 (d, J = 6.9 Hz, 2H), 7.41 (m, 4H), 7.25 (m, 1H), 7.09 (m, 2H), 6.85 (br s, 2H), 6.06 (t, J = 7.5 Hz, 1H), 2.25 (dd, J = 7.1, 6.8 Hz, 2H), 2.03 (ddd, J = 7.4, 7.1, 7.1 Hz, 2H), 1.59–1.42 (m, 4H); FDMS 513 (M+). Anal. (C<sub>30</sub>H<sub>28</sub>N<sub>2</sub>O<sub>4</sub>S·0.6CH<sub>2</sub>Cl<sub>2</sub>) C, H, N.

(Z)-7-[3-[(Benzenesulfonyl)amino] phenyl] -7- (3- pyridyl)-hept-6-enoic acid ((Z)-9c). Prepared as above in 3 steps from (Z)-13c) and benzenesulfonyl chloride (35 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.51 (dd, J = 4.6, 1.3 Hz, 1H), 8.32 (br d, J = 1.5 Hz, 1H), 7.69 (dd, J = 7.6, 1.2 Hz, 2H), 7.49-7.25 (m, 5H), 7.10 (dd, J = 7.9, 7.7 Hz, 2H), 7.02 (br d, J = 8.4 Hz, 1H), 6.85 (br d, J = 7.6 Hz, 1H), 6.79 (br d, J = 1.3 Hz, 1H), 6.05 (t, J = 7.5 Hz, 1H), 2.27 (dd, J = 7.1, 6.8 Hz, 2H), 2.04 (distorted dt, J = 7.1, 7.1 Hz, 2H), 1.60-1.42 (m, 4H); FDMS 437 (M+1).

(E)-7-[3-[(Benzenesulfonyl) amino] phenyl]-7-(3- pyridyl)-hept-6-enoic acid ((E)-9c). Prepared as above in 3 steps from methyl (E)-7-(3-nitrophenyl)-7-(3-pyridyl)-hept-6-enoate [(E)-13c] and benzenesulfonyl chloride (11 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.43 (dd, J = 4.8, 1.0 Hz, 1H), 8.39 (s, 1H), 7.74 (dd, J = 8.1, 1.1 Hz, 2H), 7.52-7.17 (m, 8H), 6.81 (br d, J = 1.4 Hz, 2H), 6.06 (t, J = 7.6 Hz, 1H), 2.28 (dd, J = 7.0, 6.8 Hz, 2H), 2.00 (dt, J = 7.2, 7.2 Hz, 2H), 1.62-1.43 (m, 4H); FDMS 437 (M+1).

(E)-7- [3- [(4-Iodobenzenesulfonyl) amino] phenyl] -7- (3-pyridyl)-hept-6-enoic acid ((E)-15g). Prepared as above in 3 steps from (E)-13c and 4-iodobenzenesulfonyl chloride (24 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.44 (br s, 2H), 7.93 (very br s, 1H), 7.72 (d, J = 8.3 Hz, 2H), 7.43 (d, J = 8.4 Hz, 2H), 7.33 (d, J = 7.9 Hz, 1H), 7.22 (m, 4H), 6.83 (buried d, 1H), 6.81 (s, 1H), 6.07 (t, J = 7.5 Hz, 1H), 2.28 (dd, J = 6.9, 6.5 Hz, 2H), 2.00 (ddd, J = 7.1, 7.1, 6.9 Hz, 2H), 1.59–1.46

(m, 4H); FDMS 563 (M+1). Anal. (C<sub>24</sub>H<sub>23</sub>IN<sub>2</sub>O<sub>4</sub>S) C, H, N

(Z)-7- [4- [(4-Iodobenzenesulfonyl) amino] phenyl] -7- (3-pyridyl)-hept-6-enoic acid (16a). Prepared as above in 3 steps from methyl (Z)-7-(4-nitrophenyl)-7-(3-pyridyl)-hept-6-enoate ((Z)-14b) (50 %):  $^{1}$  H NMR (DMSO)  $\delta$  ~10.4 (very br s, 1H), 8.48 (dd, J=3.0, 1.1 Hz, 1H), 8.23 (s, 1H), 7.88 (d, J=8.5 Hz, 2H), 7.44 (d, J=8.4 Hz, 2H), 7.38 (m, 2H), 6.96 (m, 4H), 6.09 (t, J=7.4 Hz, 1H), 2.07 (dd, J=6.9, 6.5 Hz, 2H), 1.91 (ddd, J=7.1, 6.9, 6.7 Hz, 2H), 1.37 (m, 4H); FDMS 563 (M+1). Anal. (C<sub>24</sub>H<sub>23</sub>IN<sub>2</sub>O<sub>4</sub>S) H; C: calcd, 51.25; found, 51.86; N: calcd, 4.98; found, 4.27.

(E) -8- [4- [(4-Iodobenzenesulfonyl) amino] phenyl]-8- (3-pyridyl)-oct-7-enoic acid (16b). Prepared as above in 5 steps from the ketone 12 (10%):  $^{1}$ H NMR (acetone-d<sub>6</sub>) 8 8.39 (br s, 2H), 7.92 (d, J=8.4 Hz, 2H), 7.54 (d, J=8.4 Hz, 2H), 7.48 (br d, J=8.0 Hz, 1H), 7.32 (s, 1H), 7.24 (buried, 1H), 7.23 (d, J=8.3 Hz, 2H), 7.07 (d, J=8.3 Hz, 2H), 6.16 (t, J=7.4 Hz, 1H), 2.24 (dd, J=7.3, 7.2 Hz, 2H), 2.05 (m, 2H), 1.55–1.26 (m, 6H); FDMS 577 (M+1). 96% Pure by analytical HPLC.

#### General procedure for the pyridyl ketones

4-[(tert-Butyldimethylsiloxy)methyl]phenyl 3-pyridyl ketone (18). To a mixture of 28.0 g (0.20 mol) of 1,4benzenedimethanol and 41.0 g (0.41 mol) of imidazole in ca 850 mL of anhydrous CH2Cl2 at 0 °C was cannulated 30.5 g (0.20 mol) of TBSCl in 150 mL of anhydrous CH<sub>2</sub>Cl<sub>2</sub> over a 10-15 min period. The pale yellow solution was then stirred for 3 days. The reaction was quenched with 250 mL of cold 1.0 N HCl. The organic layer was separated and washed with 250 mL of 1.0 N HCl and saturated aqueous NaHCO3 each. The aqueous layers were back-extracted with  $2 \times 500$  mL of Et<sub>2</sub>O. The combined organic layers were dried over MgSO4 and concentrated. The residue was purified by preparative HPLC using 20 % Et<sub>2</sub>O-hexanes as eluent to afford 18.60 g (36.4 %) of 4-(tert-butyldimethylsiloxy)benzyl alcohol: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  7.32 (s, 4H), 4.74 (s, 2H), 4.67 (s, 2H), 1.69 (s, 1H), 0.95 (s, 9H), 0.095 (s, 6H); MS(EI) 195 (M-t-Bu). Anal.  $(C_{14}H_{24}O_2)$  C, H.

A mixture of 18.60 g (0.074 mol) of the above benzylic alcohol and 74.4 g of MnO<sub>2</sub> in 500 mL of anhydrous THF was heated at 65–70 °C (bath temperature) for 4 h. The reaction mixture was filtered through a Celite pad and the filtrate was concentrated to yield 15.33 g (83.1 %) of clean crude 4-(tent-butyldimethylsiloxy)benzaldehyde which was used without further purification:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  9.98 (s, 1H), 7.84 (d, J = 8.0 Hz, 2H), 7.48 (d, J = 8.0 Hz, 2H), 4.80 (s, 2H), 0.94 (s, 9H), 0.10 (s, 6H); FDMS 250 (M<sup>+</sup>). Anal. (C<sub>14</sub>H<sub>22</sub> O<sub>2</sub>) C, H.

To a solution of 10.64 g (67.3 mmol) of 3-bromopyridine in ca 500 mL of anhydrous Et<sub>2</sub>O at -78 °C was added 42 mL (67.3 mmol) of 1.6 M n-BuLi in hexanes over a 35 min period. After stirring 30 min, 15.33 g (61.2 mmol) of

the above crude benzaldehyde in 100 mL of anhydrous Et<sub>2</sub>O was cannulated to the turbid lithiopyridine solution at -78 °C. After the addition, the yellow mixture was stirred for 2 h at this temperature, and then the cold bath was removed and the mixture was stirred for another 30 min. The reaction was quenched with ca 250 mL of brine and the organic layer was separated. The aqueous layer was extracted with 2 × 400 mL of CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried over MgSO<sub>4</sub>, concentrated, and purified by preparative HPLC using 2 % MeOH-CH<sub>2</sub>Cl<sub>2</sub> to afford 17.99 g (89.2 %) of the desired [4-[(tertbutyldimethylsiloxy) methyl] phenyl] (3-pyridyl) carbinol: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.61 (s, 1H), 8.46 (d, J = 3.9 Hz, 1H), 7.74 (br d, J = 7.8 Hz, 1H), 7.31 (s, 4H), 7.28 (m, 1H), 5.87 (s, 1H), 4.72 (s, 2H), 3.06 (br s, 1H), 0.93 (s, 9H), 0.087 (s, 6H); FDMS 329 (M+).

A mixture of 17.99 g (54.6 mmol) of the carbinol and 72 g of MnO<sub>2</sub> in 400 mL of THF was heated at ~65 °C (bath temperature) overnight (15 h). The oxidant was removed by filtration through a Celite pad and washed with THF and EtOAc. The combined filtrate and washing were concentrated to give 17.29 g (96.7 %) of the clean crude ketone 18 which was used without further purification:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.97 (s, 1H), 8.79 (m, 1H), 8.09 (br d, J = 7.9 Hz, 1H), 7.79 (d, J = 8.2 Hz, 2H), 7.46 (d, J = 8.4 Hz, 2H), 7.45 (buried 1H), 4.82 (s, 2H), 0.94 (s, 9H), 0.11 (s, 6H); FDMS 328 (M+1). Anal. (C<sub>19</sub>H<sub>25</sub>NO<sub>2</sub>Si) H; C: calcd, 69.68; found, 69.02; N: calcd, 4.28; found, 4.79.

3-[(tert-Butyldimethylsiloxy)methyl]phenyl 3-pyridyl ketone (17). Prepared as above from 1,3-benzenedimethanol in 4 steps (30 %):  $^{1}$ H NMR (CDCl<sub>3</sub>)  $^{3}$ 8.97 (d, J=1.7 Hz, 1H), 8.78 (dd, J=4.8, 1.5 Hz, 1H), 8.08 (dt, J=7.8, 2.0 Hz, 1H), 7.74 (s, 1H), 7.67 (d, J=7.7 Hz, 1H), 7.58 (d, J=7.7 Hz, 1H), 7.45 (m, 2H), 4.78 (s, 2H), 0.90 (s, 9H), 0.08 (s, 6H); FDMS 327 (M<sup>+</sup>). Anal. (C<sub>19</sub>H<sub>25</sub>NO<sub>2</sub>Si) C, H, N.

# General procedure for the azide intermediates

Methyl 6-(4-azidomethylphenyl)-6-(3-pyridyl)-hex-5enoates (20a). To a suspension of 3.95g (12.0 mmol) of the ketone 18 and 16.0 g (36.0 mmol) of (4carboxybutyl)triphenylphosphonium bromide in 120 mL of THF at -10 °C was added dropwise 72.0 mL (72.0 mmol) of 1.0 M t-BuOK in THF. The orange colored suspension was stirred at this temperature for 4.5 h, and the reaction was quenched with 1.0 N HCl at 0 °C while adjusting pH to ~7. The reaction mixture was concentrated to dryness, and the residue was suspended in a small amount of MeOH and filtered to remove the salt (twice). The methanolic solution was concentrated and purified by preparative HPLC using 0.5-5.0 % MeOH-CH<sub>2</sub>Cl<sub>2</sub> as eluent. The Wittig product obtained was then dissolved in 200 mL of THF and esterified with CH<sub>2</sub>N<sub>2</sub> which was generated from 1-methyl-3-nitro-1-nitrosoguanidine (5.55 g) and 5.0 N NaOH (28 mL) in 140 mL of Et<sub>2</sub>O at 0 °C. The crude reaction mixture was dried over MgSO<sub>4</sub>, concentrated, and purified by preparative HPLC with 5-10 % EtOAc-CH<sub>2</sub>Cl<sub>2</sub> to afford 2.66 g (52 %) of methyl 6-[4[(tert- butyldimethylsiloxy) methyl] phenyl]-6-(3-pyridyl)-hex-5-enoates ( $E/Z=\sim1/5$ ):  $^{1}H$  NMR (CDCl<sub>3</sub>, (Z)-isomer only)  $\delta$  8.56 (br s, 1H), 8.44 (br s, 1H), 7.49 (br d, J=7.8 Hz, 1H), 7.32 (m, 1H), 7.22 (d, J=8.2 Hz, 2H), 7.13 (d, J=8.2 Hz, 2H), 6.14 (t, J=7.5 Hz, 1H), 4.71 (s, 2H), 3.61 (s, 3H), 2.15 (dd, J=7.4, 7.4 Hz, 2H), 2.13 (ddd, J=7.6, 7.4, 7.4 Hz, 2H), 1.77 (m, 2H), 0.92 (s, 9H), 0.075 (s, 6H); FDMS 425 (M<sup>+</sup>). Anal. ( $C_{25}H_{35}NO_3Si\cdot0.4C_4H_8O_2$ ) C, H, N.

A solution of 1.13 g (2.65 mmol) of the above (E/Z)mixture (2/3) in 27 mL of THF was treated with 8.0 mL (8.0 mmol) of 1.0 M n-Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup> in THF at 0 °C for 1 h. The solvent was removed and the residue was purified by flash chromatography with 4 % MeOH-CH<sub>2</sub>Cl<sub>2</sub> as eluent to afford 718.2 mg (87.0 %) of the corresponding benzylic alcohols. To a mixture of 670 mg (2.15 mmol) of the benzylic alcohols and 0.36 mL (2.58 mmol) of Et<sub>3</sub>N in 10.75 mL of dry toluene at 0 °C was added 183 µL (2.36 mmol) of methanesulfonyl chloride, and the solution was stirred for 30 min. To this was then cannulated a solution of 1.18 g (17.2 mmol) of NaN3 and 68 mg (0.21 mmol) of n-Bu<sub>4</sub>N<sup>+</sup>Br<sup>-</sup> in 5.5 mL of H<sub>2</sub>O. The mixture was heated at 60 °C (bath temperature) for 2 h. The reaction mixture was cooled to room temperature and extracted with  $3 \times 30$ mL of EtOAc. The extract was dried over MgSO<sub>4</sub>, concentrated, and purified by flash chromatography using 2 % MeOH-CH<sub>2</sub>Cl<sub>2</sub> to afford 608.0 mg of a separable (E/Z)-mixture of azides 20a (84.1 %): <sup>1</sup>H NMR (CDCl<sub>3</sub>) (Z)-isomer only)  $\delta$  8.61 (dd, J = 4.1, 1.1 Hz, 1H), 8.48 (s, 1H), 7.70 (br d, J = 7.5 Hz, 1H), 7.51 (m, 1H), 7.24 (d, 2H), 7.16 (d, J = 8.2 Hz, 2H), 6.22 (t, J = 7.6 Hz, 1H), 4.32 (s, 2H), 3.62 (s, 3H), 2.30 (dd, J = 7.4, 7.3 Hz, 2H), 2.15(m, 2H), 1.81 (m, 2H); FDMS 336 (M+).  $(C_{19}H_{20}N_4O_2\cdot 0.2C_4H_8O_2)$  C, H, N.

Methyl 6-(3-azidomethylphenyl)-6-(3-pyridyl)-hex-5-enoates (19a). Prepared as above in 4 steps from the ketone 17 (58 %):  $^{1}$ H NMR (CDCl<sub>3</sub>, (Z)-isomer only)  $\delta$  8.57 (br d, J = 4.2 Hz, 1H), 8.44 (s, 1H), 7.46 (br d, J = 7.8 Hz, 1H), 7.33–7.10 (m, 5H), 6.16 (t, J = 7.5 Hz, 1H), 4.28 (s, 2H), 3.61 (s, 3H), 2.29 (dd, J = 7.4, 7.4 Hz, 2H), 2.15 (ddd, J = 7.6, 7.4, 7.3 Hz, 2H), 1.81 (m, 2H); FDMS 336 (M<sup>+</sup>).

General procedure for the sulfonamidomethylincorporated alkenoic acids

(Z)-6-[4-[[(Benzenesulfonyl)amino]methyl]phenyl]-6-(3-pyridyl)-hex-5-enoic acid ((Z)-22a). To a clear green solution of 158.4 mg (0.47 mmol) of the azide 20a ( $E/Z = \sim 1/8$ ) and 112.0 mg (0.47 mmol) of NiCl<sub>2</sub>·6H<sub>2</sub>O at 0 °C was added 90 mg (2.4 mmol) of NaBH<sub>4</sub> in small portions over a 20 min period. After stirring for 30 min, the mixture was concentrated and treated with 5.0 mL of cold 5.0 N HCl for 10 min. This was then basified with cold conc. NH<sub>4</sub>OH to pH 9 and extracted with 3 × 25 mL of CH<sub>2</sub>Cl<sub>2</sub>. The extract was dried over MgSO<sub>4</sub>, concentrated, and purified by flash chromatography using 5-7 % MeOH + 0.1 % conc. NH<sub>4</sub>OH in CH<sub>2</sub>Cl<sub>2</sub> to afford 83.2 mg (56.9)

%) of methyl (Z)-6-(4-aminomethylphenyl)-6-(3-pyridyl)-hex-5-enoate:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.55 (dd, J = 4.2 Hz, 1H), 8.42 (s, 1H), 7.45 (d, J = 7.8 Hz, 1H), 7.29 (m, 1H), 7.22 (d, J = 8.1 Hz, 2H), 7.14 (d, J = 8.2 Hz, 2H), 6.13 (t, J = 7.5 Hz, 1H), 3.85 (s, 2H), 3.61 (s, 3H), 2.29 (dd, J = 7.4, 7.4 Hz, 2H), 2.21 (br s, 2H), 2.13 (ddd, J = 7.5, 7.4, 7.4 Hz, 2H), 1.77 (m, 2H).

A mixture of 80.7 mg (0.26 mmol) of the above amine and 67 μL (0.52 mmol) of benzenesulfonyl chloride in 2.5 mL of pyridine was stirred at 0 °C for 4.5 h. The mixture was concentrated under reduced pressure and the residue was purified by flash chromatography using EtOAc-CH<sub>2</sub>Cl<sub>2</sub> (0-1.7) to yield 76.7 mg (65.5%) of the corresponding sulfonamide ester which was hydrolyzed with 1.0 N NaOH in THF to give, after flash chromatography with MeOH-AcOH-CH<sub>2</sub>Cl<sub>2</sub> (5:0.5:94.5), 70.7 % of the (Z)-hexenoic acid (Z)-22a: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  8.49 (br s. 1H). ~8.45 (buried, 1H), 8.34 (br s, 1H), 7.85 (d, J = 7.5 Hz, 2H), 7.55 (d, J = 7.0 Hz, 2H), 7.49 (d, J = 7.4 Hz, 2H), 7.35 (br s,1H), 7.20–7.02 (m, 4H), 6.12 (distorted t,  $J = \sim 7.1$  Hz, 1H), ~5.28 (br s, 1H), 4.10 (s, 2H), 2.32 (br s, 2H), 2.14 (m, 2H), 1.78 (br s, 2H); FDMS 436 (M<sup>+</sup>). Anal.  $(C_{24}H_{24}N_2O_4S\cdot 0.7C_2H_4O_2)$  C, H, N.

(E)-6-[4-[[(Benzenesulfonyl) amino] methyl] phenyl]-6- (3-pyridyl)-hex-5-enoic acid ((E)-22a). Prepared as above from the azide 20a ( $E/Z = \sim 8/1$ ) in 3 steps (31 %): <sup>1</sup> H NMR (CDCl<sub>3</sub>)  $\delta$  8.40 (br s, 3H), 7.86 (d, J = 7.4 Hz, 2H), 7.55-7.39 (m, 2H), 7.48 (d, J = 7.5 Hz, 2H), 7.16 (m, 3H), 7.01 (m, 2H), 6.06 (br s, 1H), 5.62 (very br s, 1H), 4.14 (s, 2H), 2.27 (br s, 2H), 2.11 (br s, 2H), 1.74 (br s, 2H); FDMS 437 (M+1). Anal. ( $C_{24}H_{24}N_2O_4S$ ) C, H, N.

(E)- and (Z)-7-[4-[[(Benzenesulfonyl)amino]methyl]-phenyl]-7-(3-pyridyl)-hept-6-enoic acids (22b). Prepared as above from the ketone 18 in 7 steps: FDMS 451 (M+1). Anal. ((E)-isomer,  $C_{25}H_{26}N_2O_4S\cdot 0.7CH_2CI_2$ ) C, H, N. 98.2 % Pure by analytical HPLC ((Z)-isomer).

(Z)-7-[4-[[(4-Iodobenzenesulfonyl)amino]methyl]phenyl]-7-(3-pyridyl)-hept-6-enoic acid (22c). Prepared as above from the ketone 18 in 7 steps: FDMS 577 (M+1). Anal.  $(C_{25}H_{25}IN_2O_4S\cdot0.6C_2H_4O_2)$  C, H, N.

(E)- and (Z)-8-[4-[[(Benzenesulfonyl)amino]methyl]-phenyl]-8-(3-pyridyl)-oct-7-enoic acids (22d). Prepared as above from the ketone 18 in 7 steps: FDMS 465 (M+1). Anal. ((Z)-isomer,  $C_{26}H_{28}N_2O_4S\cdot0.4CH_4O$ ) C, H, N.

(E)- and (Z)-6-[3-[[(Benzenesulfonyl)amino]methyl]-phenyl]-6-(3-pyridyl)-hex-5-enoic acids (21a). Prepared as above from the ketone 17 in 7 steps. (E)-isomer:  $^{1}$  H NMR (CDCl<sub>3</sub>)  $\delta$  8.40 (br s, 1H), 8.38 (br s, 1H), 7.84 (d, J = 7.1 Hz, 2H), 7.45 (m, 4H), 7.22 (m, 2H), 7.12 (br d, J = 7.8 Hz, 1H), 7.02 (s, 1H), 6.98 (d, J = 7.6 Hz, 1H), 6.09 (t, J = 7.6 Hz, 1H), 4.12 (s, 2H), 2.33 (dd, J = 6.7, 6.6 Hz, 2H), 2.13 (dt, J = 7.6, 7.5 Hz, 2H), 1.77 (m, 2H); FDMS 437 (M+1). Anal. (C<sub>24</sub>H<sub>24</sub>N<sub>2</sub>O<sub>4</sub>S) C, H, N. (Z)-isomer:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.44 (br s, 1H), 8.38 (buried, 1H), 8.32 (br s, 1H), 7.81 (d, J = 7.5 Hz, 2H), 7.56–7.42 (m, 4H), 7.33 (m, 1H), 7.11 (m, 2H), 6.99 (d, J = 0.9 Hz, 1H),

6.98 (buried, 1H), 6.08 (t, J = 7.6 Hz, 1H), 4.08 (s, 2H), 2.30 (dd, J = 7.0, 6.9 Hz, 2H), 2.13 (ddd, J = 7.5, 7.3, 7.3 Hz, 2H), 1.77 (m, 2H); FDMS 437 (M+1). Anal. ( $C_{24}H_{24}N_2O_4S \cdot 0.3C_2H_4O_2$ ) C, H, N.

- (E)- and (Z)-7-[3-[[(Benzenesulfonyl)amino]methyl]-phenyl]-7-(3-pyridyl)-hept-6-enoic acids (21b). Prepared as above from the ketone 17 in 7 steps: FDMS 451 (M+1). Anal. ( $C_{25}H_{26}N_2O_4S$ ) C, H, N.
- (E)- and (Z)-7-[3-[[(4-lodobenzenesulfonyl)amino]-methyl]phenyl]-7-(3-pyridyl)-hept-6-enoic acids (21c). Prepared as above from the ketone 17 in 7 steps. (E)-isomer:  $^1$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.43 (br s, 2H), 7.78 (d, J = 7.5 Hz, 2H), 7.54 (d, J = 8.4 Hz, 2H), 7.48 (br d, J = 8.0 Hz, 1H), 7.38-6.98 (m, 6H), 6.14 (t, J = 7.6 Hz, 1H), 4.11 (s, 2H), 2.26 (dd, J = 6.9, 6.8 Hz, 2H), 2.07 (ddd, J = 7.6, 7.5, 7.3 Hz, 2H), 1.62-1.44 (m, 4H); FDMS 577 (M+1). Anal. (C<sub>25</sub>H<sub>25</sub>IN<sub>2</sub>O<sub>4</sub>S·0.3H<sub>2</sub>O) C, H, N. (Z)-isomer:  $^1$ H NMR (CDCl<sub>3</sub>)  $\delta$  8.53 (br s, 1H), 8.36 (br s, 1H), 7.77 (d, J = 8.3 Hz, 2H), 7.47 (d, J = 8.4 Hz, 2H), 7.47 (buried, 1H), 7.35-6.98 (m, 5H), 6.08 (t, J = 7.5 Hz, 1H), 4.09 (s, 2H), 2.30 (dd, J = 7.4, 7.0 Hz, 2H), 2.10 (ddd, J = 7.3, 7.3, 7.1 Hz, 2H), 1.65-1.47 (m, 4H); FDMS 577 (M+1). Anal. (C<sub>25</sub>H<sub>25</sub>IN<sub>2</sub>O<sub>4</sub>S·0.6H<sub>2</sub>O) C, H, N.
- (E)- and (Z)-8-[3-[[(Benzenesulfonyl)amino]methyl]-phenyl]-8-(3-pyridyl)-oct-7-enoic acids (21d). Prepared as above from the ketone 17 in 7 steps: FDMS 465 (M+1). Anal. ((E/Z)-mixture:  $C_{26}H_{28}N_2O_4S\cdot 0.3C_2H_4O_2$ ; (Z)-isomer:  $C_{26}H_{28}N_2O_4S\cdot 0.6C_2H_4O_2$ ) C, H, N.

Measurement of thromboxane receptor antagonism: receptor binding assay

Membranes from outdated human platelets were prepared as previously described.<sup>25</sup> Incubations (220 µL) containing 10 µg of platelet membranes were performed in silanized glass tubes (12  $\times$  75 mm) at 30 °C for 30 min. The incubation media consisted of 10 mM Hepes, 2 mM CHAPS, 10  $\mu$ M indomethacin (pH = 7.4), ~0.05 nM (~25000 cpm) of [125I]IBOP per tube, and varying concentrations of competing ligands ranging from 10<sup>-10</sup> to 10<sup>-5</sup> M. The reaction was terminated by addition of 4 mL of ice cold buffer (25 mM Tris) at pH 7.4, followed by rapid filtration through Whatman GF/C glass filters presoaked in 0.3 % polyethyleneamine (Whatman, Inc., Clifton, NJ) using a Brandel M-24 cell harvester (Gaithersburg, MD). Non-specific binding was defined as that amount of radioactivity bound in the presence of a large molar excess (10 µM) of SQ29548, a potent TXA<sub>2</sub>/PGH<sub>2</sub> receptor antagonist.

Measurement of thromboxane synthase inhibition and prostacyclin formation

Compound or vehicle were incubated with whole human blood for 30 min at 37 °C prior to the preparation of serum as previously described. Serum  $TXB_2$  and 6-keto- $PGF_{1\alpha}$ , the stable metabolites of  $TXA_2$  and prostacyclin, respectively, were measured by radioimmunoassay as described. 27

Ex vivo experiments

Sprague Dawley rats (300 g males) were dosed by oral gavage with either vehicle (5 % acacia) or 1–10 mg/kg compound. Blood samples were collected 1 h after dosing. Animals were anesthetized (sodium pentobarbital, 87 mg/kg ip) 15 min before sample collection. Blood samples were obtained by cardiac puncture via a butterfly catheter and the first mL of blood was discarded. Blood samples were divided into duplicates and incubated at 37 °C for 1 h in 13  $\times$  100 mm glass tubes. Serum was separated by centrifugation at 2000 g for 15 min at 25 °C, transferred to polypropylene tubes, and stored at –20 °C for subsequent radioimmunoassay of TXB<sub>2</sub> produced during incubation.

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